



Causal Fungus of Side Rot on 'Zesy002' Kiwifruit in Jeju Island, South Korea

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Since the past few years, dimple rot has become one of the most serious diseases affecting *Actinidia chinensis* var. *chinensis* 'Zesy002' kiwifruit in Korea, leading to a low quality and subsequent losses of income. In the current study, the causal pathogen of side rot on 'Zesy002' kiwifruit was identified to be *Botryosphaeria dothidea*, a widespread pathogen that could penetrate the fruit directly without wounding. *Diaporthe* sp., another pathogen causing dimple rot, was isolated from infected kiwifruit. It was not able to infect fruits without wounding. Dimple-rot symptoms are known to be associated with other pathogens from the *Pseudocercospora* genus. In this study, *B. dothidea* was ratified as the causal disease agent in Jeju Island. To illustrate the pathogenicity of this fungus on 'Zesy002', further studies such as observation of infection structures of fungus may be needed.

Keywords: Diagnosis, Dimple rot, Purple blotch, Resistant cultivar, Storage disease

Introduction

In recent decades, kiwifruit production has been increasing in Asian countries including Korea. Four *Actinidia* species (*A. arguta*, *A. kolomita*, *A. polygama*, and *A. rufa*) have been grown naturally in South Korea (Shim and Ha, 1999). In the middle 1970s, the 'Hayward' variety from New Zealand was

imported to Korea. Later, a yellow-fleshed variety, *A. chinensis* var. *chinensis* cultivar 'Hort16A', was introduced to Jeju Island in 2004 (Shim and Ha, 1999). Unfortunately, a bacterial canker outbreak caused by *Pseudomonas syringae* pv. *actinidiae* (Psa) vastly occurred in orchards inside and outside New Zealand (Donati et al., 2014). In Korea, Psa was first detected on 'Hayward' variety in 1988 and on 'Hort16A' variety in 2006, resulting in an epidemic on kiwifruit orchards of south provinces, including Jeju Island (Kim et al., 2017a).

In the following years, a search for sweeter varieties with response to previous severe diseases such as canker caused by Psa led to the introduction of a new yellow-fleshed variety

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of kiwifruit, *A. chinensis* var. *chinensis* cultivar 'Zesy002', to Korea. This cultivar is grown mostly in the southern part including Jeju Island of South Korea (Zespri International Limited, 2016). However, the growing industry with this new kiwifruit cultivar has brought other challenges such as a storage disease that can reduce profits of kiwifruit farmers in South Korea. Side rot begins as a dimple and a small circular scabbed depression. It is generally developed on the side or near the stem end of the fruit, leading to darkened color from tan to dark brown. Dimples seem to appear during late fruit development. If not recognized during the harvest season, they can develop into fruit rots during cold storage, shortening shelf-life and threatening whole kiwifruit stocks (Pennycook, 1985). These symptoms were similar to those of ripe rot described by Pennycook (1985) known to develop after harvest or removal from cold storage. The causal pathogen of ripe rot is *B. dothidea* which produces lesions only for ripened kiwifruit. This fungal pathogen causes ripe rot for both kiwifruit *A. deliciosa* 'Hayward' and *A. chinensis* 'Hort16A' varieties (Manning et al., 2003). In Korea, ripe rot caused by *B. dothidea* on *A. deliciosa* 'Hayward' has also been reported (Koh et al., 2005). However, the causal pathogen of side rot on 'Zesy002' kiwifruit has not been known yet. Because side rot occurs not only in late stage, but also in early stage of fruits, the causal pathogen could be different from that causing ripe rot. Furthermore, it has been reported that *Pseudocercospora hangzhouensis* can cause side or dimple rot symptom on *A. chinensis* 'Hort16A' in Japan and Korea (Yano et al., 2015). Therefore, the exact diagnosis of pathogen on 'Zesy002' kiwifruit causing side rot may be necessary.

In the present study, a diagnosis was carried out for infected 'Zesy002' kiwifruit showing side rot in Jeju orchards. Furthermore, pathogenicities of *B. dothidea* isolates on 'Zesy002' kiwifruit cultivars with or without mechanical wounding were investigated. Similarly, to find the causal fungus on side rot, pathogenicities of both fungi *B. dothidea* and *Pseudocercospora* sp. were determined by inoculation them onto 'Zesy002' kiwifruit.

Materials and Methods

Isolation of pathogenic fungi and morphological diagnosis. A total of 64 'Zesy002' kiwifruit showing dimple rot (Fig. 1) or purple blotch were collected from three orchards in Jeju Island of Korea during the 2018 harvest season. These

fruits were surface-sterilized by dipping in 1% (v/v) sodium hypochlorite for 60 sec followed by 70% (v/v) ethanol for 60 sec, rinsed twice with sterile distilled water for 60 sec, and air-dried for 1 hr. Kiwifruit skin was peeled back and four segments (1 cm^2) were detached from margins of symptomatic underlying tissues and transferred to potato dextrose agar (PDA) plates containing 100 $\mu\text{g/ml}$ of streptomycin sulphate (Sigma Aldrich, St. Louis, MO, USA). After incubation at 28°C for 6 days, representative fungal hyphae from each plate were transferred to oatmeal agar medium. For sporulation of the fungus, plates were incubated under light with air supply at 28°C for 7 days. Morphological characteristics of fungi were observed under a light microscope (Olympus, Tokyo, Japan).

Molecular identification. Eight isolates from dimple rot and two from purple blotch symptom were selected for molecular identification. Genomic DNA was extracted from approximately 200 mg of fungal hyphae scraped directly from PDA-grown cultures using the cetyltrimethylammonium bromide method (Aamir et al., 2015) with slight modifications. Briefly, hyphae were lyophilized in liquid nitrogen and grinded with mortar and pestle. The powder obtained was placed in a 2 μl vial. Then 700 μl of lysis buffer (100 mM Tris HCl [pH 8.0], 50 mM EDTA, 3% sodium dodecyl sulfate) was added. This mixture was homogenized with a WiseMix Multifunction Vortex Mixer (DAIHAN Scientific, Wonju, Korea) for 20 min and centrifuged at 13,000 rpm for 10 min. The supernatant was transferred to a fresh 2 ml microcentrifuge tube. Then 2 μl of RNase A (10 mg/ml) was added and the whole solution was incubated at 37°C for 15 min. Later, according to the volume in each vial, equal volume of cleaning buffer (phenol: chloroform: Isoamyl alcohol, 25:24:1) was added. The mixture was homogenized and centrifuged at 13,000 rpm for 10 min. This step was repeated once. The upper aqueous layer was taken in a fresh 1.5 ml micro centrifuge tube and equal volume of 100% ethanol was added. DNA in this solution was precipitated at -20°C for 30 min followed by centrifugation at 12,000 rpm for 10 min. The pellet of DNA was washed with 70% ethanol and centrifuged again at 12,000 rpm for 5 min. Finally, the DNA pellet was air-dried overnight and re-suspended in 1× TE buffer.

For each sample, 40 μl of PCR amplification cocktail contained 2 μl of DNA template (5–10 ng/ml), 10 pmol of each forward and reverse primer, 25 mM of MgCl₂, 2.5 mM of each

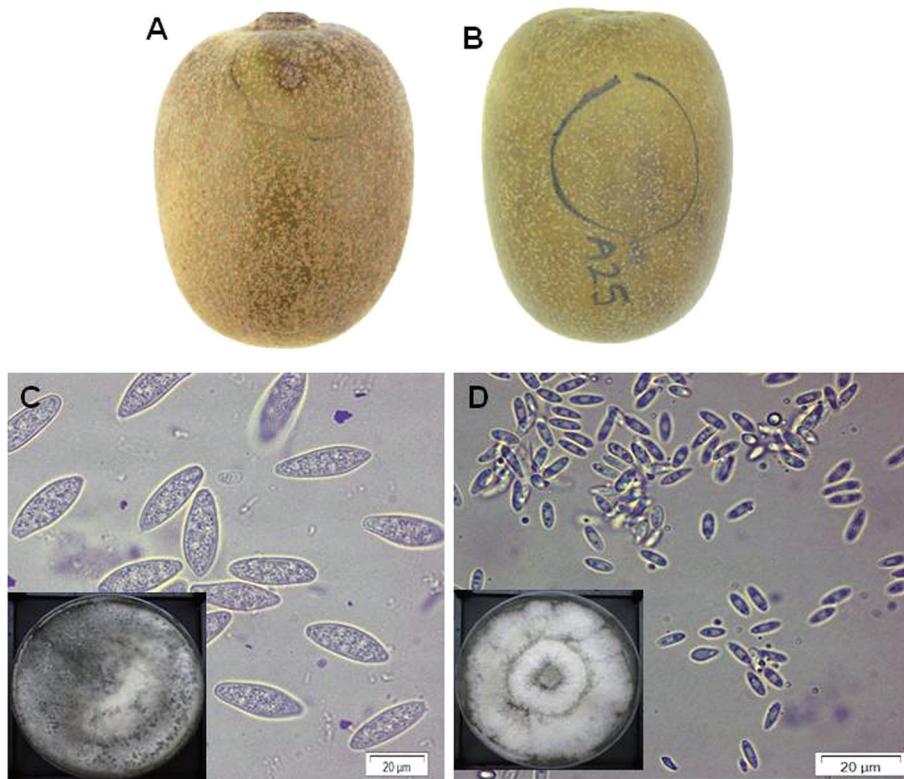


Fig. 1. Typical symptom of dimple rot (A) and purple blotch (B) on 'Zesy002' kiwifruit collected from the orchards in Jeju Island, South Korea during 2018 harvest season. Morphological characteristics of the conidia of isolate C1.1 for type I (C) and A24.3 for type II (D) grown oatmeal agar under light and air supply condition. The square boxes were plates from which the conidia were harvested. The presented photos were taken at 7 days after inoculation. Scale bars=20 µm (C, D).

of the four dNTPs, and 5 units of Taq DNA polymerase. Internal transcribed spacer (ITS) rDNA region was amplified for all selected isolates using universal primers ITS1 (5'-TCCGTAG-GTGAACCTGCGG-3') and 4 (5'-TCCTCCGCTTATTGATAT-GC-3') (White et al., 1990). A set of *B. dothidea* specific primers of FAF (5'-CATCCGCAGCGTGGGAGAACAT-3') and Bt2b (5'-ACCCTCAGTGTAGTGACCCTGGC-3') were used to re-identify the fungus (Xu et al., 2016). The reaction for ITS was performed on a thermal cycler (Biometra TOne 96G, Analytik Jena AG, Jena, Germany). PCR cycling parameters were: an initial preheat at 94°C for 5 min, followed by 35 cycles of denaturation at 94°C for 45 sec, annealing for 45 sec at 58°C, and extension at 72°C for 1 min, with a final extension stage at 72°C for 10 min. In the case of using primer set of FAF/Bt2b, the annealing temperature was set at 60°C.

PCR products were detected by 1% agarose gel electrophoresis and stained with 0.01% ethidium bromide for UV light visualization using an UVP transilluminator (UVsolo touch, Analytik Jena AG). Nucleotide sequences of PCR

products were determined using an ABI 3730xl DNA Analyzer (Macrogen Inc., Seoul, Korea). Resulting sequences were compared with sequences in GenBank database using BLAST program of NCBI (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) for primary identification.

Pathogenicity test of both isolates and other pathogens. Asymptomatic mature 'Zesy002' kiwifruit were obtained from Zespri-Korea. These kiwifruits were surface-sterilized by dipping in 1% (v/v) sodium hypochlorite for 60 sec followed by 70% (v/v) ethanol for 60 sec, rinsed twice with sterile distilled water for 60 sec, and air-dried for 1 hr. Unwounded and wounded fruits were used. For preparing wounded fruits, kiwifruit were punctured 2–3 mm deep on the upper part with a sterile wood stick right before fungal inoculation.

Representative cultures of both fungal types (isolate C1.1 for Type I and isolate A24.3 for type II) (Supplementary Table 1) were used as inoculum. Several *Pseudocercospora* species

have been previously reported to be commonly associated with this kind of side-rot disorder in other kiwifruit varieties (Yano et al., 2015). Thus, *Pseudocercospora* sp. isolate obtained from the Department of Plant Medicine of Sunchon National University was added as an inoculum. These fungi were transferred into PDA medium and incubated under aerobic conditions with 5,000 lux at 28°C for 7 days. Agar plugs with a size of 5 mm in diameter were detached from the PDA medium and inoculated onto wounded and unwounded kiwifruit. PDA plugs without fungus were used as negative controls. These inoculated fruits were placed in baskets and incubated at 28°C with 95% humidity under no light for 7 days. These experiments were repeated three times with temporal separation. Three replications were carried out in every experiment.

Data analysis. For all experiments, data were analyzed with Duncan's multiple range test using statistical analysis

system (SAS) program (version 9.0, SAS Institute Inc., Cary, NC, USA). Statistical significance was considered at $P < 0.05$.

Results

Isolation of pathogenic fungi and morphological diagnosis. A total number of 64 isolates from 61 fruits were obtained (Supplementary Table 1). Among them, most fungal isolates showed phenotypical characteristics similar to those of the main phenotype "Type I" presented principally in kiwifruit with dimple rot symptoms (Fig. 1A). The other phenotype "Type II" was less frequent, although it was also recurrent in kiwifruit with purple blotch symptoms (Fig. 1B).

The conidia of isolate type I showed morphological characteristics similar to those of *B. dothidea* described in other kiwifruit cultivars (Tyson, 2018; Zhou et al., 2015). Furthermore, this fungus has been reported to be the major pathogen associated with postharvest rot on kiwifruit in Korea (Koh et

Table 1. Identification of isolates from different orchards in Jeju Island by BLAST

| Isolates | Symptom | Primer | Accession no. | Description | Identity (%) |
|----------|---------------|--------|---------------|--------------------------------|--------------|
| A2.1 | Dimple | ITS4 | KX610122.1 | <i>Botryosphaeria dothidea</i> | 98.3 |
| | | FAF | KX306959.1 | <i>B. dothidea</i> | 100 |
| A3.2 | Dimple | ITS4 | JX179253.1 | <i>B. dothidea</i> | 98.6 |
| | | FAF | HQ529720.1 | <i>B. dothidea</i> | 100 |
| A4.2 | Dimple | ITS4 | JQ936677.1 | <i>B. dothidea</i> | 100 |
| | | FAF | KP183132.1 | <i>B. dothidea</i> | 100 |
| A13.4 | Dimple | ITS4 | JX179253.1 | <i>B. dothidea</i> | 99.7 |
| | | FAF | KP183132.1 | <i>B. dothidea</i> | 99.6 |
| B6.2 | Dimple | ITS4 | JX179253.1 | <i>B. dothidea</i> | 96.3 |
| | | FAF | KP183130.1 | <i>B. dothidea</i> | 98.3 |
| C1.1 | Dimple | ITS4 | JX179253.1 | <i>B. dothidea</i> | 99.1 |
| | | FAF | KP183132.1 | <i>B. dothidea</i> | 99.3 |
| C19.1 | Dimple | ITS4 | JQ936677.1 | <i>B. dothidea</i> | 99.0 |
| | | FAF | KP183129.1 | <i>B. dothidea</i> | 99.0 |
| A15.1 | Dimple | ITS4 | KC145875.1 | <i>Diaporthe</i> sp. | 98.8 |
| | | FAF | - | nb | - |
| A22.1 | Purple blotch | ITS4 | KJ609017.1 | <i>D. phaseolorum</i> | 99.1 |
| | | FAF | - | nb | - |
| A24.3 | Purple blotch | ITS4 | KC145874.1 | <i>Diaporthe</i> sp. | 99.1 |
| | | FAF | - | nb | - |

nb, no band by PCR.

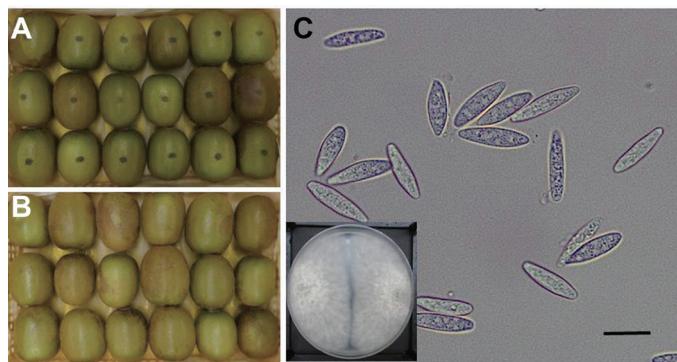


Fig. 2. Side rot symptom on the 'Zesy002' kiwifruit after artificially inoculated with the conidial suspension of isolate I *B. dothidea* with concentration of 1×10^5 conidia/ml (A). On the negative control treated with sterilized H₂O showed no symptom (B). The presented photos were taken at 7 days after inoculation. Morphological characteristics of the fungal conidia re-isolated from infected kiwifruit (C) which was same with those of first isolated fungus. The presented small square in the down was plate incubated of the re-isolate on PDA for 7 days. Scale bar=20 μm (C).

al., 2003; Kwon et al., 2011). Based on these morphological observations, this type I isolate was suspected as *B. dothidea* whose conidia were oval to fusoid in shape, aseptate, measuring 7 ± 1 μm in width and 34 ± 3 μm in length. Morphological characteristics of another isolate type II were similar to those of *Diaporthe citri* (Tyson, 2018; Zhou et al., 2015) whose α-conidia were oval to fusoid in shape, aseptate, measuring 3 ± 1 μm in width and 7 ± 2 μm in length. They have also been found on harvested kiwifruit in Korea (Koh et al., 2003; Kwon et al., 2011).

Molecular identification. Total DNAs were extracted from all 10 selected isolates for PCR. By using ITS primers, total DNAs of all isolates causing dimple rot and purple blotch were amplified. After sequencing, BLASTn showed that all isolates causing dimple rot shared 96–100% sequence identities with *B. dothidea* strains except for isolate A15.1 (Table 1). Two isolates A22.1 and A24.3 causing purple blotch were identified as *D. phaseolorum* and *Diaporthe* sp., respectively (Table 1). These results nearly coincided with results of morphological identification (Table 1, Fig. 2, Supplementary Table 1). Using a specific primer set of FAF and Bt2b for *B. dothidea*, DNA sequences of isolates causing dimple rot were amplified to have a size of 322 bp, but not isolates causing purple blotch (Table 1, Fig. 3). As expected, BLASTn analysis

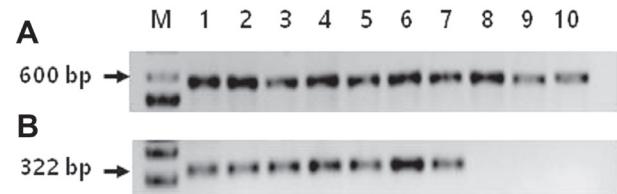


Fig. 3. Gel electrophoresis of PCR-amplified internal transcript spacer (ITS) regions from the isolates using universal primers (A) and using specific primers for *B. dothidea* FAF and Bt2b (B). M, 1 kb ladder (iNtRON Bio); lane 1 (A2.1), lane 2 (A3.2), lane 3 (A4.2), lane 4 (A13.4), lane 5 (B6.2), lane 6 (C1.1), lane 7 (C7.2), and lane 8 (A15.1) were isolates from dimple rot and lane 9 (A22.1) and lane 10 (A24.3) from purple blotch.

revealed that the amplified gene shared 98–100% sequence identities with *B. dothidea* (Table 1).

Pathogenicity tests of both isolates and other pathogens. Typical symptoms of dimple rot were observed for wounded 'Zesy002' kiwifruit inoculated with type I isolate *B. dothidea* (Fig. 4A). Unexpectedly, no purple blotch was observed. On the other hand, symptoms like dimple rot were found for wounded kiwifruit inoculated with type II isolate *Diaporthe* sp. (Fig. 4B). However, *Pseudocercospora* sp. did not cause any symptom on wounded kiwifruit (Fig. 4C), indicating that 'Zesy002' kiwifruit might not be a host of this fungus. On wounded fruits without fungal inoculation, no symptoms were found (Fig. 4D).

Type I isolate *B. dothidea* successfully infected unwounded kiwifruit (Fig. 4E), indicating their pathogenicity to Gold 3 kiwifruit in nature. However, type II isolate *Diaporthe* sp. could not infect kiwifruit (Fig. 4F), indicated that type II isolate could not infect Gold 3 kiwifruit by itself. Similarly, symptoms were not observed on unwounded 'Zesy002' kiwifruit inoculated with *Pseudocercospora* sp. (Fig. 4G), meaning that *Pseudocercospora* sp. might not be the major pathogen for dimple rot. On unwounded kiwifruit, no symptom was found (Fig. 4H).

Discussion

Postharvest dimple rot constitutes a serious challenge to kiwifruit production in Jeju Island, Korea. Pathogenic fungi associated with this disease on 'Zesy002' kiwifruit need to be identified in order to decrease economical loss of kiwifruit farms. Dimple-like and purple blotch lesions were recognized (Whiteman, 2018) in orchard or storage during 2018–2019

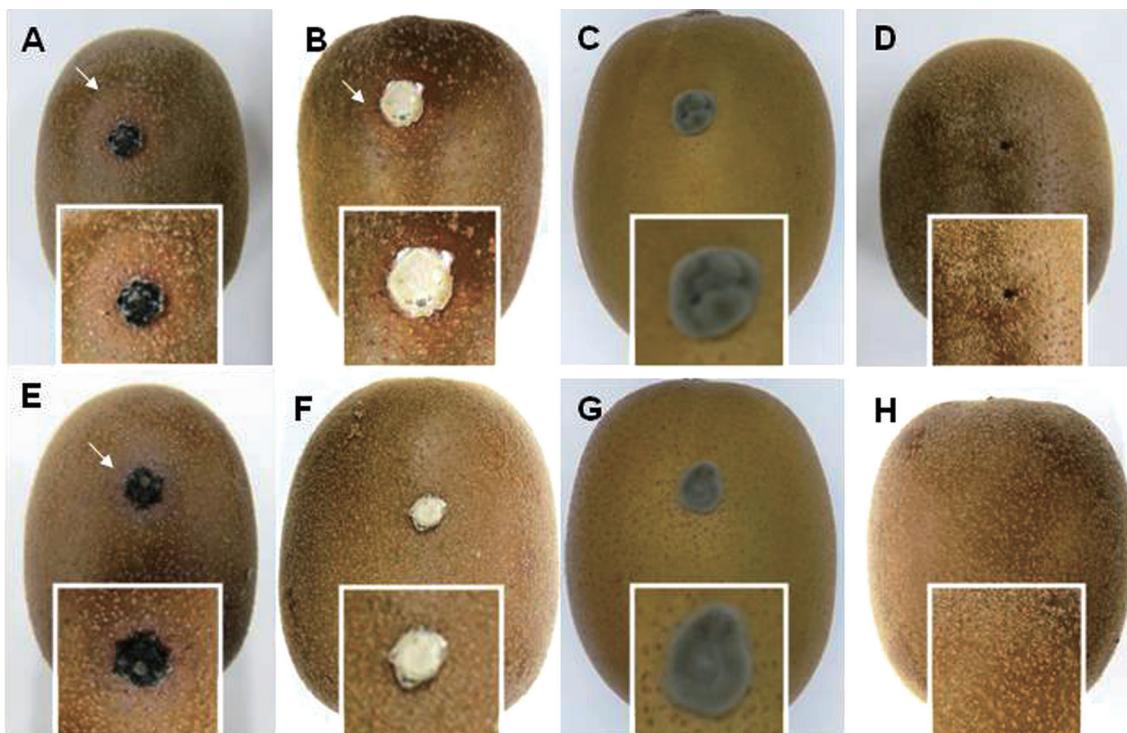


Fig. 4. External symptoms of dimple rot on artificially inoculated on the wounded/unwounded 'Zesy002' kiwifruit with the type I isolate *B. dothidea* (A, E), type II isolate *Diaporthe* sp. (B, F), *Pseudocercospora* sp. (C, G), and sterile agar plug (D, H), respectively. The presented photos were taken at 7 days after inoculation. The square boxes represented the magnified of the inoculated part. The arrow showed the symptom of dimple rot.

harvest seasons on 150 'Zesy002' kiwifruit from 13 greenhouses around Jeju Island. Dimple symptoms cause small scabbed depressions on the side or near the stem end of the kiwifruit. They can develop internally into a side rot. Purple blotch symptoms cause darker areas of the fruit of a purplish color. In some cases, the fruit skin looks dried and shrunken. Identical dimple signs have been reported in several other kiwifruit cultivars in Korea, China, New Zealand, and other countries (Kim et al., 2017b; Koh et al., 2003; Kwon et al., 2011; Manning et al., 2003; Zhou et al., 2015), but not in 'Zesy002' kiwifruit. In the present work, these symptoms were excised and cultured, resulting in 64 isolates, from which 95% of them showed similar morphological characteristics (Fig. 2). Isolate type I from kiwifruit showed morphological characteristics similar to those of fungus on other kiwifruit cultivars described before. It was identified as *Botryosphaeria dothidea* (Tyson, 2018; Zhou et al., 2015). Their abundant gray to olive aerial mycelia and the hyaline unicellular α -conidia of oval shape matched with the Ascomycota genus, *Botryosphaeria*. DNA sequence identification of a sample of 50 isolates shared 100% identities with *B. dothidea* strains (Fig. 3). Fur-

thermore, this fungus has been reported to be the major pathogen associated with postharvest rot on kiwifruit in Korea with incidence of 83.3% to 95.4% (Koh et al., 2003; Kwon et al., 2011). Based on our inoculation experiments and morphological observations, this type I isolate was identified as *B. dothidea*.

Isolate type II seemed to be *Diaporthe citri* reported on harvest kiwifruit with a frequency of 4.6% to 11.9% in Korea (Koh et al., 2003; Kwon et al., 2011). It has been reported that this fungus could not infect kiwifruit without wound (Li et al. 2017), similar to finding of the present study. This result suggests that *D. citri* may be saprophytic in kiwifruit. It needs wound by pathogenic fungus such as *B. dothidea* to infect kiwifruit.

B. dothidea is a globally widespread pathogen associated with dieback, canker, and severe fruit rots. It has an extensive host range (Beckman and Reilly, 2005; Milholland, 1972; Pitt et al., 2010; Tang et al., 2012). Through pathogenicity testing, it was found that *B. dothidea* could penetrate the fruit directly without wounding. However, the presence of a wound exacerbates this disease, accelerating its development and

intensifying its severity (Li et al., 2017; Zhou et al., 2015). After re-isolation from artificially inoculated kiwifruit, the same symptoms in the fruit as well as the same cultural behavior, morphology, and DNA sequence identity were found (Fig. 3), thus confirming Koch's Postulates and ratifying *B. dothidea* as a causal agent of postharvest dimple rot on 'Zesy002' kiwifruit in Jeju Island.

In South Korea, yellow-fleshed varieties have shown to be less resistant to postharvest rots than green fleshed ones (Kwon et al., 2011). Ploidy level might play a fundamental role in breeding resistance against bacterial canker caused by *Psa* (Tahir et al., 2019), a pathogenic bacterium that can lead to severe crop loss. Therefore, evaluation of susceptibilities to *B. dothidea* among 'Hort16A' (diploid), 'Zesy002' (tetraploid) and 'Hayward' (hexaploid) varieties may provide insights about a possible relationship between predisposition to dimple rot and ploidy level. However, dimple-rot susceptibility might not follow the ploidy hypothesis applied for *Psa*. Additional research is needed to find candidate alleles for susceptibility and resistance to dimple-rot disease.

On the other hand, solitary dimples on the side of kiwifruit have been previously reported as characteristic symptoms caused by *Mycosphaerellaceae* pathogens *Pseudocercospora actinidiae* and *Pseudocercospora hangzhouensis* (Yano et al., 2015). This possibility was explored by comparing the pathogenicity capacity of *Pseudocercospora* sp. with that of *B. dothidea* under the same conditions. While *B. dothidea* was able to penetrate the unwounded fruit skin, this strain of *Pseudocercospora* sp. did not result in any symptoms. It was not re-isolated from any inoculated kiwifruit, indicating that *Pseudocercospora* sp. might not be the major pathogen for this form of dimple rot in 'Zesy002'. These results open new possibilities for integral management of dimple rot disease, in which protecting the fruit during crucial developmental stages and designing different storage conditions might better complement control strategies for 'Zesy002' kiwifruit disease. Further experimentation regarding *B. dothidea* life cycle, overwintering of secondary inoculum, and the possibility of symbiotic relations with fruit feeding insects may be needed to develop a successful strategy for 'Zesy002' kiwifruit protection in Jeju Island.

Conflicts of Interest

The current study was financially supported by Zespri Group

Limited. Sonia Whiteman is employee of Zespri International Limited. All other authors have no competing interests.

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Electronic Supplementary Material

Supplementary materials are available at Research in Plant Disease website (<http://www.online-rpd.org/>).

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